# POSITRON EMISSION TOMOGRAPHY AS A TOOL FOR STUDYING ALCOHOL ABUSE

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ositron emission tomography (PET) is an imaging technology that measures the concentration, distribution, and pharmacokinetics of radiotracers—molecules that are labeled with short-lived positron-emitting variants (i.e., radioisotopes) of chemical elements naturally found in the body. These radioisotopes can be attached to compounds involved in normal brain function and then injected into the blood stream. For example, radioactive carbon-11 (11C) and fluorine-18 (18F) can be used to label the sugar glucose, which is the brain's only energy source, and oxygen-15 (15O) can be used to label water molecules, which can help measure blood flow in the brain. The signals emitted by these radiotracers then are measured using specific detectors. For example, for brain measurements, detectors arranged in a ring around the subject's head collect the data, which are then transferred to a computer and converted into a three-dimensional image of the brain. Because these measurements are noninvasive, the technology allows researchers to track biochemical transformations in the living human and animal body. PET is a highly sensitive method; it measures radioisotope concentrations in the nanomolar to picomolar range (10<sup>-19</sup> to 10<sup>-12</sup> M) (Schmidt 2002). Therefore, the technique can be used to label compounds that are of pharmacological and physiological relevance. These radiotracers then can be used to probe neurochemical and metabolic processes at the relevant physiological concentrations without perturbing the system that is measured.

To exert their effects on the brain, alcohol and other drugs (AODs) act on signaling molecules (i.e., neurotransmitters) in the brain as well as on the molecules on the surface of neurons (i.e., receptors) with which the neurotransmitters interact. (For more information on nerve signal transmission, neurotransmitters, and their receptors, see the article by Lovinger, pp. 196–214.) Specific compounds that selectively bind to such receptors, to the molecules that transport neurotransmitters back into cells, and to the enzymes that are involved in the synthesis or metabolism of neurotransmitters can be labeled for use as PET radiotracers. As a result, PET can be used to assess the metabolic and neurochemical actions of AODs and to evaluate the consequences of chronic AOD use (Volkow et al. 1997; Wang et al. 2000; Wong et al. 2003). Since its

inception, PET has been used extensively to study the effects of AODs in human and nonhuman primates; however, the recent development of microPET technology has expanded its applications to research in rodents. In addition, increasing numbers of studies are using PET methodology to assess the involvement of genetic variations in individual genes (i.e., polymorphisms) in brain function and neurochemistry. This article specifically summarizes the role of PET as a tool for alcohol neuroscience research. The studies discussed are divided into those that assess the effects of alcohol on brain function (i.e., brain metabolism and cerebral blood flow) and those that assess its effects on neurochemistry.

### PET Analyses of Brain Function

Indicators of brain function, such as cerebral blood flow, glucose utilization, and oxygen consumption, are the most common signals detected in functional brain-imaging techniques. These metabolic signals have been examined in a variety of disorders, primarily through the use of (18F)-fluoro-2deoxyglucose (FDG) as a radiotracer in PET imaging. Thirty-two years after its introduction, FDG still is the most widely used radiopharmaceutical for PET studies. This type of PET imaging allows the noninvasive observation of glucose utilization by different types of brain cells, including neurons and supporting cells known as glial cells (Magistretti and Pellerin 1996). In the brain, the sugar glucose is metabolized to lactate, which is a preferred energy source for neurons. Accordingly, glucose metabolism is a powerful indicator of brain function. FDG-PET imaging has the potential to detect very early brain dysfunction, even before neuropsychological testing yields abnormal results. In addition, the technique can be used to monitor treatment response and the effects of possible therapeutic intervention against the disease.

PET analyses using FDG to measure brain glucose metabolism and radiolabeled water to measure cerebral blood flow have been used to study the acute and chronic effects of alcohol in nonalcoholic control subjects, alcoholics, and people at risk of alcoholism (e.g., children of

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alcoholics). Other PET studies using FDG have examined alcohol's toxic effects on neurons (i.e., neurotoxicity) or gender-specific responses to alcohol. The findings include the following:

- Acute alcohol administration markedly reduced brain glucose metabolism throughout the whole brain, including the prefrontal cortex (Volkow et al. 2006) (see figure 1), whereas it increases cerebral blood flow in some brain regions, such as the prefrontal cortex (Volkow et al. 2007). In addition, it was shown that alcoholics displayed both a prefrontal modulation (i.e., reduced brain glucose) in the activity of cells using the neurotransmitter dopamine, combined with a profound decrease in dopamine activity (Volkow et al. 2007). These data suggested that interventions to restore prefrontal regulation and the dopamine deficit could be therapeutically beneficial in alcoholics (Volkow et al. 2007). Moreover, normally, brain metabolism and cerebral blood flow are coupled—that is, areas that show high brain metabolism also exhibit high blood flow and vice versa. Thus, these findings also suggest that alcohol dissociates this metabolic flow coupling.
- A recent FDG–PET study demonstrated abnormally low function of a brain region called the thalamus, which processes and relays information from other brain regions, in alcoholics suffering from acute alcohol-related hallucinations (Soyka et al. 2005).
- Alcoholics and normal subjects respond differently to an acute alcohol challenge, with the alcoholics showing a smaller behavioral response but larger decrease in brain metabolism than normal subjects (Volkow et al. 1993).
- Regional brain metabolic changes in response to treatment with the benzodiazepine medication lorazepam, which, like alcohol, enhances the activity of the neurotransmitter γ-aminobutyric acid (GABA), differed between alcoholic and control subjects. The findings likely indicate altered function of a certain type of GABA receptor (i.e., the GABA–BZ receptor) in alcoholics (Volkow et al. 1995). Indeed, the pattern of regional brain metabolic decrements seen with acute alcohol administration is similar to that observed after acute administration of lorazepam in healthy people, supporting the hypothesis that alcohol and benzodiazepines have a common molecular target for some metabolic effects (Wang et al. 2000).
- Studies measuring brain glucose metabolism or cerebral blood flow documented reduced activity in frontal and parietal cortical regions in alcoholics. This observation is consistent with findings from neuropsychological studies showing that alcoholics have deficits in executive function and attention, which are controlled by these brain areas. Overall, these studies strongly support the concept that alcoholism is associated with damage to the frontal and parietal lobes.

 Several studies have used imaging to probe the recovery of brain function after alcohol withdrawal. These studies found that the alcohol-related decreases in brain glucose metabolism partially recover in abstinent alcoholics, particularly during the first 16 to 30 days after withdrawal (Volkow et al. 1994).

Imaging studies also have addressed the influence of gender on the effects of alcoholism on the brain. It generally is believed that women are more vulnerable to alcohol's toxic effects than men. However, whereas male alcoholics have consistently shown reductions in brain glucose metabolism relative to control subjects, a PET study using <sup>18</sup>FDG in 10 recently detoxified female alcoholics reported no differences between alcoholics and control females (Wang et al. 1998). These results do not support the assumption that alcohol has greater toxic effects on the female brain, at least with respect to regional brain glucose metabolism. However, it should be noted that the severity of alcohol use in these female alcoholics was less than that of the male alcoholics previously investigated in PET studies. Therefore, studies in male subjects with moderately severe alcoholism are required to confirm gender differences in sensitivity to alcohol's effects on brain metabolism.

# PET Analyses of Neurotransmitters and Receptor Binding

PET imaging also has been an effective tool in examining neurotransmitter systems associated with alcohol abuse and alcoholism (for a review of the various neurotransmitter systems affected by alcohol, see Koob 2003; Koob and Le Moal 2008). PET studies have shown that several neurotransmitters appear to mediate alcohol's reinforcing and addictive effects (Wang et al. 2000). Of these, dopamine is believed to play perhaps the most important role in mediating alcohol's reinforcing effects by acting on a brain circuit called the mesolimbic dopamine system¹ (Fowler and Volkow 1998). Researchers have used a plethora of radiolabeled compounds to examine various components of the dopamine system using PET analyses, including the following:

- [11C]m-tyrosine, a radiolabeled variant of the amino acid tyrosine, which is the starting material for dopamine synthesis;
- [18F]DOPA, a radiolabeled variant of a compound known as 3,4-dihydroxy-L-phenylalanine (L-DOPA), which is an intermediate product in dopamine synthesis;
- A molecule called [¹C]DTBZ (dihydroytetrabenzine), which helps measure the activity of the vesicular monoamine transporter (VMAT)—a transport protein that helps

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<sup>&</sup>lt;sup>1</sup> This brain circuit primarily involves two brain regions called the ventral tegmental area (VTA) and the nucleus accumbens (NAc). It plays a central role in reward, motivation, and reinforcement. Its activity also is controlled by certain areas of the prefrontal cortex.

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transport dopamine and other signaling molecules into the vesicles in which they are stored in the signal-emitting (i.e., presynaptic) neuron;

- [11C]cocaine, which helps measure the activity of the dopamine transporter (DAT) that shuttles released dopamine back into the presynaptic cells;
- A compound called [¹C]SCH23390 that helps determine the activity of a certain dopamine receptor, the D<sub>1</sub> dopamine receptor (D<sub>1</sub>R); and
- A molecule called [¹C]raclopride, which helps measure the activity of another dopamine receptor, the D<sub>2</sub> dopamine receptor (D<sub>2</sub>R).

PET imaging studies, as well as postmortem studies of alcoholic subjects, have indicated that D<sub>2</sub>R levels may be involved with alcohol addiction, because the levels of these receptors were reduced in the striatum of the brains of alcoholic subjects (Heinz et al. 2004; Volkow et al. 1996). Additional PET analyses using [<sup>11</sup>C]raclopride demonstrated that higher D<sub>2</sub>R availability in nonalcoholic members of alcoholic families may protect these individuals against alcoholism (Volkow et al. 2006). The data also supported the notion that the low D<sub>2</sub>R levels observed in alcoholics may reflect the effects of chronic alcohol exposures.

Other PET studies have used [ $^{11}$ C]raclopride to assess changes in dopamine induced by stimulant drugs as a measure of the reactivity of dopamine-releasing cells. This approach is based on the fact that [ $^{11}$ C]raclopride competes for binding to  $D_2$  receptors with endogenous dopamine—

that is, the more endogenous dopamine is released by the neurons, the less [11C]raclopride can bind to the receptor and vice versa. Thus, changes in specific [11C]raclopride binding that occur after stimulant administration reflect the relative increases in dopamine induced by the drug. Several studies have revealed a decrease in dopamine release in alcoholic subjects, particularly in the ventral striatum (Martinez et al. 2005; Volkow et al. 2007). In contrast, clinical studies comparing people with a positive family history for alcoholism and people without such a family history did not show differences between the two groups in stimulant-induced dopamine increases in the striatum. These data suggest that the decreased dopamine release in alcoholics is caused by chronic alcohol exposure (Monro et al. 2006). The investigators postulated that the decreased reactivity of the mesolimbic dopamine system in alcoholics could put them at risk of consuming large amounts of alcohol to compensate for deficiencies in this reward pathway.

Investigators also have conducted PET studies using multiple tracers simultaneously to study the relationship between the changes in dopamine activity (as assessed with [11C]raclopride) and brain glucose metabolism in the prefrontal cortex (as measured with FDG). These studies demonstrated a negative association between brain glucose metabolism in prefrontal cortical regions (i.e., cingulated gyrus, dorsolateral cortex, and orbitofrontal cortex) and changes in dopamine levels in the striatum (which also contains the nucleus accumbens) of control subjects. Thus, the higher the metabolism in the prefrontal region the lower the changes in dopamine levels. In alcoholic subjects, in contrast, the activity in the prefrontal cortical regions was not correlated with dopamine changes in

the striatum (Volkow et al. 2007). These findings suggest that in alcoholics the normal regulation of dopamine cell activity by signals from the prefrontal cortex is disrupted; thus, the decreased dopamine cell activity in alcoholics may represent abnormal prefrontal regulation of the mesolimbic dopamine system.

Another study measured the activity of the vesicular monoamine transporters in alcoholics. This study, which used a radiotracer specific for one type of these transporters (i.e., [11C]DTB2), revealed that the levels of this transporter were reduced in the striatum, suggesting that the damaging effects of severe chronic alcoholism on the central nervous system are more extensive than previously considered (Gilman et al. 1998).

PET imaging studies also have been used to examine the role of neurotransmitters known as endogenous opioids in alcohol dependence. Studies

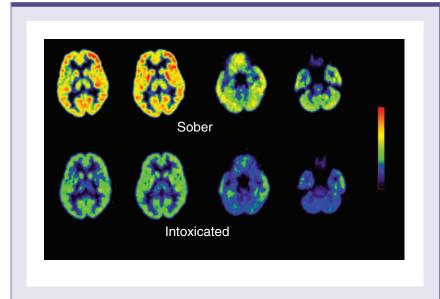


Figure 1 Brain activity during alcohol intoxication. Alcohol drinking markedly reduces brain metabolism.

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using a radiolabeled synthetic opioid pain reliever,  $[^{11}C]$  carfentanil, showed that the severity of alcohol craving correlated with an increase in a certain type of opiate receptor (i.e., the  $\mu$ -opiate receptors) in the ventral striatum and, particularly, the nucleus accumbens (Heinz et al. 2005). These findings point to a neuronal correlate of the alcohol craving observed in abstinent alcoholic patients.

Finally, PET analyses have helped examine the neuro-chemistry underlying the relationship between alcoholism and aggression and, more specifically, whether signal transmission mediated by the neurotransmitter serotonin contributes to this relationship (Brown et al. 2007). The investigators evaluated the density of the serotonin transporter in alcoholic patients who were assessed for aggressive characteristics. The results showed that none of the clinical measures used, including measures of aggression, correlated with serotonin transporter binding in the alcoholic subjects.

## **FUTURE DIRECTIONS**

The studies reviewed here reflect the potential of PET as a tool to investigate the alcoholic brain. However, there are additional opportunities for using this technology to investigate the neural underpinnings of alcoholism. For example, PET can be applied to examine the consequences of genetic variations (i.e., polymorphisms), gene modifications, or stem cell procedures on regional brain function in alcoholics.

Other studies have demonstrated the feasibility of using PET to investigate the role of genes in the rodent brain. This development has extended the usefulness of PET in elucidating the role of genes in brain function, aging, and adaptations to environmental and pharmacologic interventions for alcoholism. Technologies to completely disrupt (i.e., "knock out") or newly introduce (i.e., "knock in") certain genes in mice have been particularly valuable in elucidating the role of genes and the proteins they encode in normal and pathological behaviors (Avale et al. 2004; Gainetdinov and Caron 2000; Gainetdinov et al. 2001). Other technological advances, such as smallanimal PET imaging and microPET technology have rapidly progressed since their introduction (Cherry 1997) and today offer PET images with a resolution of just under 2 mm. Furthermore, the combination of microPET images with images of the same animals obtained using other technologies (e.g., high-field magnetic resonance imaging) has allowed researchers to extend the use of PET imaging studies to rodent models of psychiatric disease (Ding et al. 2004; Rodriguez-Gomez et al. 2007; Thanos et al. 2002, 2008a,b,c,d). Thus, microPET has become an effective in vivo imaging tool for noninvasively studying rodent models of alcohol abuse.

#### FINANCIAL DISCLOSURE

The authors declare that they have no competing financial interests.

#### REFERENCES

AVALE, M.E.; FALZONE, T.L.; GELMAN, D.M.; ET AL. The dopamine D4 receptor is essential for hyperactivity and impaired behavioral inhibition in a mouse model of attention deficit/hyperactivity disorder. *Molecular Psychiatry* 9(7):718–726, 2004. PMID: 14699433

Brown, A.K.; George, D.T.; Fujita, M.; et al. PET [11C]DASB imaging of serotonin transporters in patients with alcoholism. *Alcoholism: Clinical and Experimental Research* 31(1):28–32, 2007. PMID: 17207098

CHERRY, S.R. MicroPET: A high resolution PET scanner for imaging small animals. *IEEE Transactions on Nuclear Science* 44:1161–1166, 1997.

DING, Y.S.; GATLEY, S.J.; THANOS, P.K.; ET AL. Brain kinetics of methylphenidate (Ritalin) enantiomers after oral administration. *Synapse* 53(3):168–175, 2004. PMID: 15236349

FOWLER, J.S., AND VOLKOW, N.D. PET imaging studies in drug abuse. *Journal of Toxicology. Clinical Toxicology* 36(3):163–174, 1998. PMID: 9656971

GAINETDINOV, R.R., AND CARON, M.G. An animal model of attention deficit hyperactivity disorder. *Molecular Medicine Today* 6:43–44, 2000.

GAINETDINOV, R.R.; MOHN, A.R.; AND CARON, M.G. Genetic animal models: Focus on schizophrenia. *Trends in Neuroscience* 24(9):527–533, 2001. PMID: 11506886

GILMAN, S.; KOEPPE, R.A.; JUNCK, L.; ET AL. Benzodiazepine receptor binding in cerebellar degenerations studied with positron emission tomography. *Annals of Neurology* 38(2):176–185, 1995. PMID: 7654065

GILMAN, S.; KOEPPE, R.A.; ADAMS, K.M.; ET AL. Decreased striatal monoaminergic terminals in severe chronic alcoholism demonstrated with (+)[11C]dihydrotetrabenazine and positron emission tomography. *Annals of Neurology* 44(3):326–333, 1998. PMID: 9749598

HEINZ, A.; SIESSMEIER, T.; WRASE, J.; ET AL. Correlation between dopamine D2 receptors in the ventral striatum and central processing of alcohol cues and craving. *American Journal of Psychiatry* 161(10):1783–1789, 2004. PMID: 15465974

HEINZ, A.; REIMOLD, M.; WRASE, J.; ET AL. Correlation of stable elevations in striatal mu-opioid receptor availability in detoxified alcoholic patients with alcohol craving: A positron emission tomography study using carbon 11-labeled carfentanil. *Archives of General Psychiatry* 62(1):57–64, 2005. PMID: 15630073

KOOB G.F. Alcoholism: Allostasis and beyond. *Alcoholism: Clinical and Experimental Research* 27(2):232–243. 2003. PMID: 12605072

KOOB G.F., AND LE MOAL, M. Addiction and the brain antireward system. Annual Review of Psychology 59:29–53, 2008. PMID: 18154498

MAGISTRETTI, P.J., AND PELLERIN, L. The contribution of astrocytes to the 18F-2-deoxyglucose signal in PET activation studies. *Molecular Psychiatry* 1(6):445–452, 1996. PMID: 9154245

MARTINEZ, D.; GIL, R.; SLIFSTEIN, M.; ET AL. Alcohol dependence is associated with blunted dopamine transmission in the ventral striatum. *Biological Psychiatry* 58(10):779–786, 2005. PMID: 16018986

MCBRIDE, W.J.; CHERNET, E.; DYR, W.; ET AL. Densities of dopamine D2 receptors are reduced in CNS regions of alcohol-preferring P rats. *Alcohol* 10(5):387–390, 1993. PMID: 8216885

PHILLIPS, T.J.; BROWN, K.J.; BURKHART-KASCH, S.; ET AL. Alcohol preference and sensitivity are markedly reduced in mice lacking dopamine D2 receptors. *Nature Neuroscience* 1(7):610–615, 1998. PMID: 10196569

RODRIGUEZ-GOMEZ, J.A.; LU, J.Q.; VELASCO, I.; ET AL. Persistent dopamine functions of neurons derived from embryonic stem cells in a rodent model of Parkinson's disease. *Stem Cells* 25(4):918–928, 2007. PMID: 17170065

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SCHMIDT, K.C., AND TURKHEIMER, F.E. Kinetic modeling in positron emission tomography. *Quarterly Journal of Nuclear Medicine* 46(1):70–85, 2002. PMID: 12072847

SOYKA, M.; KOCH, W.; AND TATSCH, K. Thalamic hypofunction in alcohol hallucinosis: FDG PET findings. *Psychiatry Research* 139(3):259–262, 2005. PMID: 16043330

STEFANINI, E.; FRAU, M.; GARAU, M.G.; ET AL. Alcohol-preferring rats have fewer dopamine D2 receptors in the limbic system. *Alcohol and Alcoholism* 27(2):127–130, 1992. PMID: 1355967

THANOS, P.K.; VOLKOW, N.D.; FREIMUTH, P.; ET AL. Overexpression of dopamine D2 receptors reduces alcohol self-administration. *Journal of Neurochemistry* 78(5):1094–1103, 2001. PMID: 11553683

THANOS, P.K.; TAINTOR, N.B.; ALEXOFF, D.; ET AL. In vivo comparative imaging of dopamine D2 knockout and wild-type mice with (11)C-raclopride and microPET. *Journal of Nuclear Medicine* 43(11):1570-1577, 2002. PMID: 12411561

THANOS, P.K.; TAINTOR, N.B.; RIVERA, S.N.; ET AL. DRD2 gene transfer into the nucleus accumbens core of the alcohol preferring and nonpreferring rats attenuates alcohol drinking. *Alcoholism: Clinical and Experimental Research* 28(5):720–728, 2004. PMID: 15166646

THANOS, P.K.; RIVERA, S.N.; WEAVER, K.; ET AL. Dopamine D2R DNA transfer in dopamine D2 receptor-deficient mice: Effects on ethanol drinking. *Life Sciences* 77(2):130–139, 2005. PMID: 15862598

THANOS, P.K.; MICHAELIDES, M.; PIYIS, Y.K.; ET AL. Food restriction markedly increases dopamine D2 receptor (D2R) in a rat model of obesity as assessed with in-vivo muPET imaging ([(11)C] raclopride) and in-vitro ([3H] spiperone) autoradiography. *Synapse* 62(1):50–61, 2008a. PMID: 17960763

THANOS, P.K.; MICHAELIDES, M.; BENVENISTE, H.; ET AL. The effects of cocaine on regional brain glucose metabolism is attenuated in dopamine transporter knockout mice. *Synapse* 62(5):319–324, 2008*b*. PMID: 18286542

THANOS, P.K.; MICHAELIDES, M.; GISPERT, J.D.; ET AL. Differences in response to food stimuli in a rat model of obesity: In-vivo assessment of brain glucose metabolism. *International Journal of Obesity (London)* 32(7):1171–1179, 2008c. PMID: 18475275

THANOS, P.K.; HWANG, Y.F.; PATEL, U.; ET AL. "In-Vivo Changes in Brain Glucose Metabolism and Dopamine D2 Receptor Availability in Response to Acute Ethanol Binge Drinking in Rats." Presented at the 2nd International Conference on Applications of Neuroimaging to Alcoholism (ICANA 2), New Haven, CT, January 19–20 2008d.

VOLKOW, N.D.; WANG, G.J.; HITZEMANN, R.; ET AL. Decreased cerebral response to inhibitory neurotransmission in alcoholics. *American Journal of Psychiatry* 150(3):417–422, 1993. PMID: 8382010

VOLKOW, N.D.; WANG, G.J.; HITZEMANN, R.; ET AL. Recovery of brain glucose metabolism in detoxified alcoholics. *American Journal of Psychiatry* 151(2):178–183, 1994. PMID: 8296885

VOLKOW, N.D.; WANG, G.J.; BEGLETTER, H.; ET AL. Regional brain metabolic response to lorazepam in subjects at risk for alcoholism. *Alcoholism: Clinical and Experimental Research* 19(2):510–516, 1995. PMID: 7625590

VOLKOW, N.D.; WANG, G.J.; FOWLER, J.S.; ET AL. Decreases in dopamine receptors but not in dopamine transporters in alcoholics. *Alcoholism: Clinical and Experimental Research* 20(9):1594–1598, 1996. PMID: 8986209

VOLKOW, N.D.; WANG, G.J.; OVERALL, J.E.; ET AL. Regional brain metabolic response to lorazepam in alcoholics during early and late alcohol detoxification. *Alcoholism: Clinical and Experimental Research* 21(7):1278–1284, 1997. PMID: 9347090

VOLKOW, N.D.; WANG, G.J.; FOWLER, J.S.; ET AL. Cocaine abusers show a blunted response to alcohol intoxication in limbic brain regions. *Life Sciences* 66(12):PL161–167, 2000. PMID: 10737368

VOLKOW, N.D.; WANG, G.J.; FRANCESCHI, D.; ET AL. Low doses of alcohol substantially decrease glucose metabolism in the human brain. *Neuroimage* 29(1):295–301, 2006*a*. PMID: 16085426

VOLKOW, N.D.; WANG, G.J.; BEGLEITER, H.; ET AL. High levels of dopamine D2 receptors in unaffected members of alcoholic families: Possible protective factors. *Archives of General Psychiatry*. 63(9):999–1008, 2006*b*. PMID: 16953002

VOLKOW, N.D.; WANG, G.J.; TELANG, F.; ET AL. Profound decreases in dopamine release in striatum in detoxified alcoholics: Possible orbitofrontal involvement. *Journal of Neuroscience* 27(46):12700–12706, 2007. PMID: 18003850

WANG, G.J.; VOLKOW, N.D.; FOWLER, J.S.; ET AL. Regional cerebral metabolism in female alcoholics of moderate severity does not differ from that of controls. *Alcoholism: Clinical and Experimental Research* 22(8):1850–1854, 1998. PMID: 9835307

WANG, G.J.; VOLKOW, N.D.; FRANCESCHI, D.; ET AL. Regional brain metabolism during alcohol intoxication. *Alcoholism: Clinical and Experimental Research* 24(6):822–829, 2000. PMID: 10888070

Wong, D.F.; Maini, A.; Rousset, O.G.; and Brasic, J.R. Positron emission tomography: A tool for identifying the effects of alcohol dependence on the brain. *Alcohol Research & Health* 27(2):161–173, 2003. PMID: 15303627

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